

TRUFORMA[®] Point-of-Care Canine **Cortisol Assay**

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Key Messages

- Accurate and precise measurement of cortisol levels in canines is needed to diagnose and manage hypoadrenocorticism and hyperadrenocorticism.
- The TRUFORMA platform uses innovative bulk acoustic wave (BAW) technology to provide a nonoptical and fluorescence-free detection system for diagnostic use at the point of care (POC) in veterinary clinics.
- The dynamic range of the TRUFORMA canine cortisol assay allows the quantification of high and low concentrations of cortisol within the same assay, which is vital for accurate diagnosis of hyperadrenocorticism and hypoadrenocorticism in canines.
- The high precision and correlation to a reference laboratory assay shown for the TRUFORMA canine cortisol assay provides veterinarians with accurate and reliable diagnostic results at the POC, providing opportunities for improved patient treatment and real-time client communication.



Introduction

Accurately diagnosing canine adrenal dysfunction is challenging in veterinary practice due to the complexity of current reference lab methodologies for immunoassays and the variability in performance of the POC tests. There is a need for a POC cortisol assay that has been optimized for canine samples. The TRUFORMA platform uses BAW sensor technology to provide veterinarians rapid, reliable, and accurate measurement of canine cortisol levels at the POC.

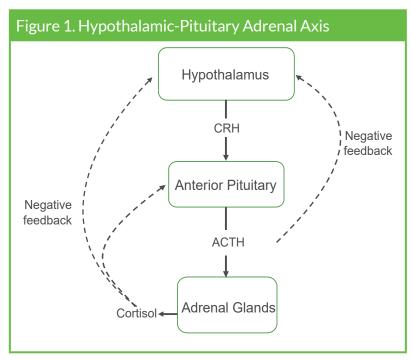
The objectives of this study were to:

- Determine analytical performance attributes for the TRUFORMA cortisol assay.
- Describe how the TRUFORMA canine assay differs from currently available assay.
- Compare TRUFORMA canine assay performance with an assay used as part of the standard of care at veterinary diagnostic laboratories.

Clinical Significance of Canine Cortisol Testing

Cortisol is a steroid hormone produced by the adrenal glands in response to adrenocorticotropic hormone (ACTH) from the pituitary gland. Cortisol release occurs in response to stimulation — including stress, hypoglycemia,

and physical exercise - and is regulated through a negative feedback loop (Figure 1). Two major adrenal disorders include hyperadrenocorticism, also known as Cushing's disease, and hypoadrenocorticism, also known as Addison's disease. Hyperadrenocorticism is caused by overproduction of adrenal hormones and is most commonly the result of pituitary or adrenal gland tumors or excessive use of steroids. Hypoadrenocorticism is caused by decreased adrenal hormone production and is most often due to immune-mediated destruction of the adrenal glands. Other causes of hypoadrenocorticism include adrenal gland destruction from physical trauma or infection, pituitary or adrenal tumors, abrupt withdrawal from steroid treatment, or excessive use of adrenolytic agents.



CRH, corticotrophin-releasing hormone; ACTH, adrenocorticotropic hormone



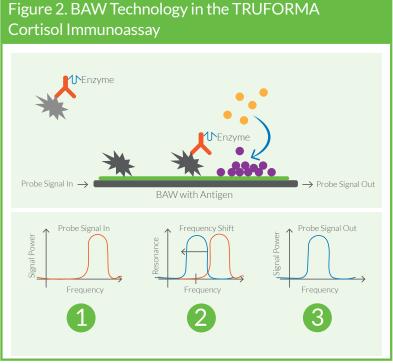
Canine Cortisol

A definitive diagnosis of adrenal disease in canines currently relies on comparing serum cortisol levels at baseline to levels post-dynamic adrenal testing. Dynamic adrenal testing for diagnosing and differentiating hyperadrenocorticism includes ACTH stimulation testing, low-dose dexamethasone suppression testing (LDDST), and/or high-dose dexamethasone suppression testing (HDDST). ¹ Measurement of basal cortisol can also be used as a screening test to rule out or exclude the diagnosis of hypoadrenocorticism.^{2,3,4} Once a diagnosis of hyperadrenocorticism has been reached, a patient's baseline cortisol as well as levels post-ACTH stimulation aid in optimizing treatment dosages and long-term monitoring of patients. This is especially useful for avoiding iatrogenic adrenocortical atrophy in Cushing's patients. Several assays — including radioimmunoassay (RIA), chemiluminescent assay, and enzyme-linked immunosorbent assay (ELISA) — are available for measuring cortisol levels at reference labs and have been validated for use in canines.⁵ However, the limited dynamic range of many of these cortisol assays, especially suboptimal lower limits of quantification, increases the risk for misdiagnosis of hypoadrenocorticism. A limited number of commercial POC cortisol assays are also available, but validation data from these assays have not been published in peer-reviewed literature.¹ Clearly, there is a need for accurate, reliable quantification of cortisol at the POC.

- Accurate and reliable measurements of cortisol levels are needed for the diagnosis of adrenal dysfunction in canines.
- A high-performance veterinary POC immunoassay for cortisol, with published performance data, is needed.

TRUFORMA Platform

The TRUFORMA platform uses BAW sensor technology to provide a nonoptical and fluorescence-free detection system for diagnostic use at the POC in clinics. BAW technology is extremely reliable and precise, and has been well tested in products across industries such as telecommunications and aerospace. Functionalized BAW biosensors consist of multiple resonators, each composed of a piezoelectric material subjected to an electrical field. The resonators can be coated with detection reagents, such as antibodies and nucleic acids, for immunoassay and molecular testing. Whereas current enzyme-based immunoassays use optical sensors to detect the generation of luminescence or color change, BAW biosensors used as part of TRUFORMA assays measure both binding events and the insoluble product that is generated by the enzymes that accumulate on the sensor surface, thereby



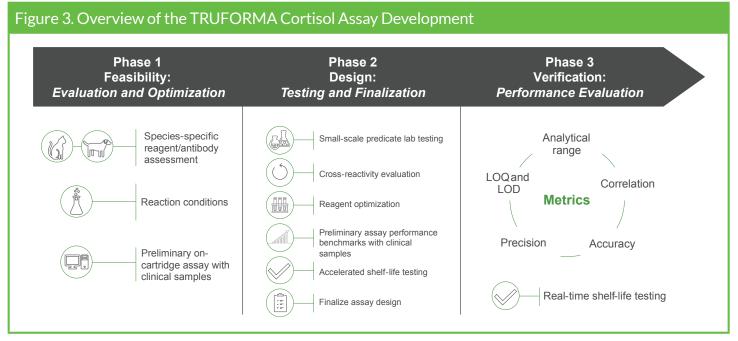
The TRUFORMA cortisol assay is a competitive immunoassay in which the BAW sensor is coated with antigen (dark gray). Antigen present in the sample (light gray) binds to an antibody-enzyme conjugate in solution and prevents the antibody from binding to the antigen-coated biosensor. After several wash steps, an enzyme substrate is exposed to the BAW biosensor surface and bound enzyme converts the substrate to an insoluble product that accumulates on the BAW biosensor surface. This is measured as a shift in frequency by the BAW biosensor. The signal is proportional to the amount of analyte present in the sample.



creating a frequency shift in resonance proportional to the mass accumulated on the sensor (Figure 2). Veterinary medical professionals will be the first to use the BAW sensor technology in a POC diagnostic setting due to human diagnostic test premarket approval timelines.

Cortisol Assay Development Overview

The TRUFORMA cortisol assay is a competitive immunoassay that uses a monoclonal anti-cortisol antibody, selected for optimal performance for canine testing. Using the industry standard recommendations for bioanalytical method validation and the Clinical and Laboratory Standards Institute (CLSI) guidelines on method comparison and bias estimation,⁶ the TRUFORMA assay performance requirements were chosen to meet or exceed reference laboratory performance in order to provide unparalleled performance at the POC. The three phases of the canine cortisol assay development were designed to provide a high-quality and reliable POC assay and included feasibility evaluation and optimization with species-specific assessment, design and testing of preliminary assay performance, and performance verification (Figure 3).



LOD, limit of detection; LOQ, limit of quantitation

Assay Verification Results

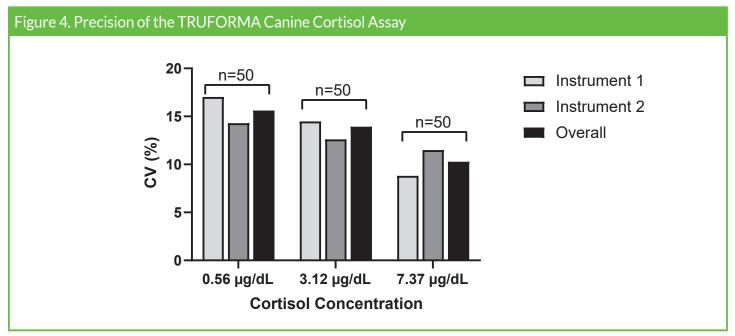
The TRUFORMA canine cortisol assay analytical performance was evaluated and compared with the reference-laboratory predicate Siemens IMMULITE[®] 2000 cortisol assay, an automated solid-phase chemiluminescent competitive immunoassay. Samples from healthy and diseased canines as well as spiked canine samples were analyzed. Baseline cortisol samples and samples collected after ACTH stimulation were also included.



Analytical Precision

Analytical precision was evaluated by measuring the variability in assay results (between-run percentage coefficient of variation [%CV]) under the normal operating conditions in the laboratory. Precision was evaluated by testing three canine samples with varying cortisol concentrations. Each sample was tested with five replicates over five separate days on two different instruments, for a total of 150 results. Observed %CV was calculated.

The TRUFORMA canine cortisol assay demonstrated an overall %CV of <20% (Figure 4). For each sample, the overall %CV was comparable to the %CV for each instrument, indicating repeatability across instruments. A \pm 25% between-runs %CV is recommended and is considered a quality %CV for measuring assay precision in a ligand-binding assay.⁷



%CV was calculated for three serum samples with varying cortisol concentrations using 150 runs. One statisical outlier was removed from the sample at 3.12 µg/dL following CLSI EP05-A3 guidelines. CV, coefficient of variation.

Time to Test Results (TTR), Dynamic Range, and Limit of Quantitation

Time to Test Results (TTR) refers to the amount of time for TRUFORMA to initiate the test and generate the result. The POC testing of the TRUFORMA platform provides more timely and actionable results, which can improve patient care through faster clinical decision making and communication with pet owners.

Dynamic range refers to the span of test result values that can be accurately measured, and limit of quantitation (LOQ) refers to the lowest (lower LOQ [LLOQ]) and highest (upper LOQ [ULOQ]) analyte concentrations that can be reliably detected with predefined accuracy and precision.

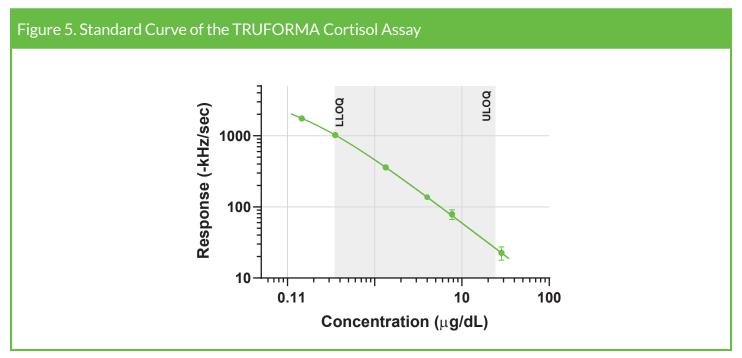
The TRUFORMA cortisol assay had an LLOQ that was slightly lower than that of the IMMULITE Cortisol assay (Table 1). The TRUFORMA cortisol assay's dynamic range allows the quantification of both clinically high and clinically low cortisol concentrations. The enhanced dynamic range can improve the ability to diagnose and treat at the POC without the need to send samples to a reference laboratory.



Table 1. Summary of TTR, Dynamic Range and LLOQ for the TRUFORMA Canine Cortisol Assay Compared With the Siemens IMMULITE Cortisol Assay		
Test	TRUFORMA	IMMULITE
	Canine Optimized	Human Optimized
TTR, minutes	<16.0	>30.0
Dynamic range, µg/dL	0.349-24.0	1.0-50.0
LLOQ, µg/dL	0.349	1.0

LLOQ, lower limit of quantitation; TTR, time-to-result.

Six calibrators with known concentrations of cortisol were tested using the TRUFORMA cortisol cartridge. Each calibrator was run with nine replicates across three different instruments, and the average value was used to generate a standard curve. The linearity and reportable range of the TRUFORMA cortisol assay illustrates linear performance within the clinically relevant range (Figure 5).



Six calibrators with known concentrations of cortisol were used to generate a standard curve for canine analysis. The shaded region represents the reported dynamic range. LLOQ, lower limit of quantitation; ULOQ, upper limit of quantitation.

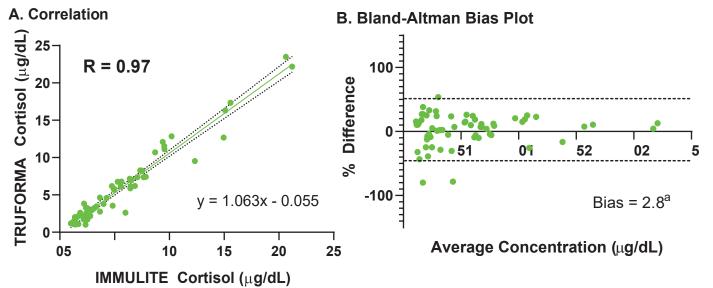


Assay Correlation Between TRUFORMA Cortisol Assay and Siemens IMMULITE Canine Cortisol Assay

Assay correlation and Bland-Altman bias plot analysis evaluate the agreement and commutability of a new test method with a comparative or reference method. A total of 61 serum samples were run on the same freeze-thaw cycle on the TRUFORMA and Siemens IMMULITE devices. The instrument reports concentrations based on the standard curve, and results were used to generate correlation and bias plots.

The TRUFORMA canine cortisol assay showed high correlation (R=0.97) with the Siemens IMMULITE (Figure 6A), with no apparent bias (mean bias =2.8%; 95% CI, -3.4 to 9.0%) (Figure 6B) across the dynamic range.





(A) Correlation studies were performed comparing the results from the TRUFORMA canine cortisol and IMMULITE cortisol assays using 61 canine samples. Dotted lines represent 95% CI for the linear regression line. (B) Bland-Altman bias plots were generated by plotting the mean concentrations vs. the % difference ([TRUFORMA – IMMULITE]/mean). Dotted lines represent 95% limits of agreement. "The 95% CI for the bias includes the line of equality, indicating no bias.



Cross-Reactivity

Known amounts of cortisol, and potential cross-reactants were added to depleted serum and tested in triplicate using the TRUFORMA cortisol assay.

Cross-reactivity was measured in the TRUFORMA cortisol assay, and the results are reported in Table 2.

Table 2. Summary of Cross-Reactivity for the TRUFORMA Cortisol Assay		
Material	Concentration, µg/dL	Cross-Reactivity, %
Aldosterone	1000	ND
Triamcinolone	5000	ND
Spironolactone	1000	ND
Estriol	100	ND
Estrone	500	ND
Dexamethasone	400	5.7
Methylprednisolone	200	4.0
Corticosterone	400	2.3
Cortisone	400	0.4
Progesterone	400	0.2
Betamethasone	1000	0.2
Methotrexate	100	0.2
17 α-hyroxyprogesterone	400	0.1
Desoxycorticosterone pivalate	1000	0.1
	100	0.1
	10	ND
Trilostane	1000	0.1
	100	ND
	10	ND
Prednisolone	240	13.1
	8	ND
Prednisone	5000	0.2
	16	ND

ND, not detected



Conclusions

The TRUFORMA canine cortisol assay demonstrated high precision with a wide dynamic range, providing confidence in the reliability of cortisol results at the POC. The TRUFORMA canine cortisol assay's dynamic range allows the quantification of both high and low cortisol concentrations within the same assay, which is vital for diagnosing adrenal disease in canines. The availability of the TRUFORMA cortisol assay optimized for canine samples at the POC allows for immediate testing of baseline and dynamic adrenal samples, leading to timely diagnosis and immediate initiation of lifelong treatment. Additionally, the improved LLOQ will increase confidence in the diagnosis of hypoadrenocorticism and interpretation of LDDST and HDDST results compared to the currently available commercial assays.

The TRUFORMA canine cortisol assay provides veterinarians with accurate and reliable diagnostic results at the POC, allowing for a more rapid and informed patient diagnosis, and improved treatment and client communication.



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Abbreviations and Acronyms

- ACTH Adrenocorticotropic Hormone
- BAW Bulk Acoustic Wave
- CLSI Clinical and Laboratory Standards Institute
- CRH Corticotrophin-Releasing Hormone
- CV Coefficient of Variation
- ELISA Enzyme-Linked Immunosorbent Assay
- HDDST High-Dose Dexamethasone Suppression Test
- LDDST Low-Dose Dexamethasone Suppression Test
- LOD Limit of Detection
- LOQ Limit of Quantitation
- LLOQ Lower Limit of Quantitation
- ND Not Detected
- POC Point of Care
- RIA Radioimmunoassay
- TTR Time to Test Results
- ULOQ Upper Limit of Quantitation

References

- 1. Feldman E, Nelson R, Reusch C, Scott-Moncrieff JC. Canine and Feline Endocrinology. 4th ed. Philadelphia, PA: WB Saunders; 2014.
- 2. Bovens C, Tennant K, Reeve J, Murphy KF. Basal Serum Cortisol Concentration as a Screening Test for Hypoadrenocorticism in Dogs. J Vet Intern Med. 2014.28(5):1541-5.
- 3. Gold AJ, Lanlois DK, Refsal KR. Evaluation of basal serum or plasma cortisol concentrations for the diagnosis of hypoadrenocorticism in dogs. J Vet Intern Med. 2016. 30: 1798-1805.
- 4. Lennon EM, Boyle TE, Hutchins EG, et al. Use of basal serum or plasma cortisol concentrations to rule out a diagnosis of hypoadrenocorticism in dogs: 123 cases (2000-2005). J Am Vet Med Assoc. 2007. 231: 413-416.
- 5. Russell NJ, Foster S, Clark P, et al. Comparison of radioimmunoassay and chemiluminescent assay methods to estimate canine blood cortisol concentrations. Aust Vet J. 2007. 85:487-94.
- 6. Clinical and Laboratory Standards Institute. EP09-A3: Measurement Procedure and Bias Estimation Using Patient Samples. Wayne, PA. 2018.
- 7. US Food and Drug Administration. Bioanalytical method validation. 2018.